

# *Model of $\alpha$ -linolenic acid metabolism*

***N.Kokulan, C.-H. Lai***  
***School of Computing and Mathematical Sciences***  
***University of Greenwich***  
***London, UK***

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# OVERVIEW

*Introduction*

*compartmental model of  $\alpha$ -linolenic acid metabolism*

*The set of differential equations*

*Parameters estimation*

*Numerical Results*

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## Introduction

- *A mathematical model of  $\alpha$ -linolenic acid metabolism is derived from the experimental data of plasma concentration [1] at different times for  $18:3n-3$ ,  $20:5n-3$ ,  $22:5n-3$  and  $22:6n-3$  in 95 healthy volunteers who have been ingested with 1g of isotope tracer of  $\alpha$ -linolenic acid.*
- *Mathematical methods based on inverse problems for the identification of unknown kinetics parameters in the system of ODEs using time series measurements.*
- *The goal of this model is to obtain kinetics parameters which can be used to quantify the biosynthesis of long chain  $n-3$  PUFA beginning with  $\alpha$ -linolenic acid.*

## *Compartmental model of $\alpha$ -linolenic acid metabolism*

- *A physiological compartmental model of  $\alpha$ -linolenic acid metabolism is employed with well-known consideration of metabolic pathway for  $\alpha$ -linolenic.*

### **Assumptions**

- *The liver is the main site for the biosynthesis of fatty acid.*
- *As a liver biopsy was not possible in these subjects therefore in this study rate constants determined from the model represents kinetics of fatty acid from their plasma pool concentrations.*
- *The model presumes that each rate constant reflects several steps of the metabolism that occur within the liver [1]*
- *It is assumed that this kinetic function is similar to the appearance of all the fatty acids measured in the plasma.*

# Compartmental model for the metabolism of $\alpha$ -linolenic acid

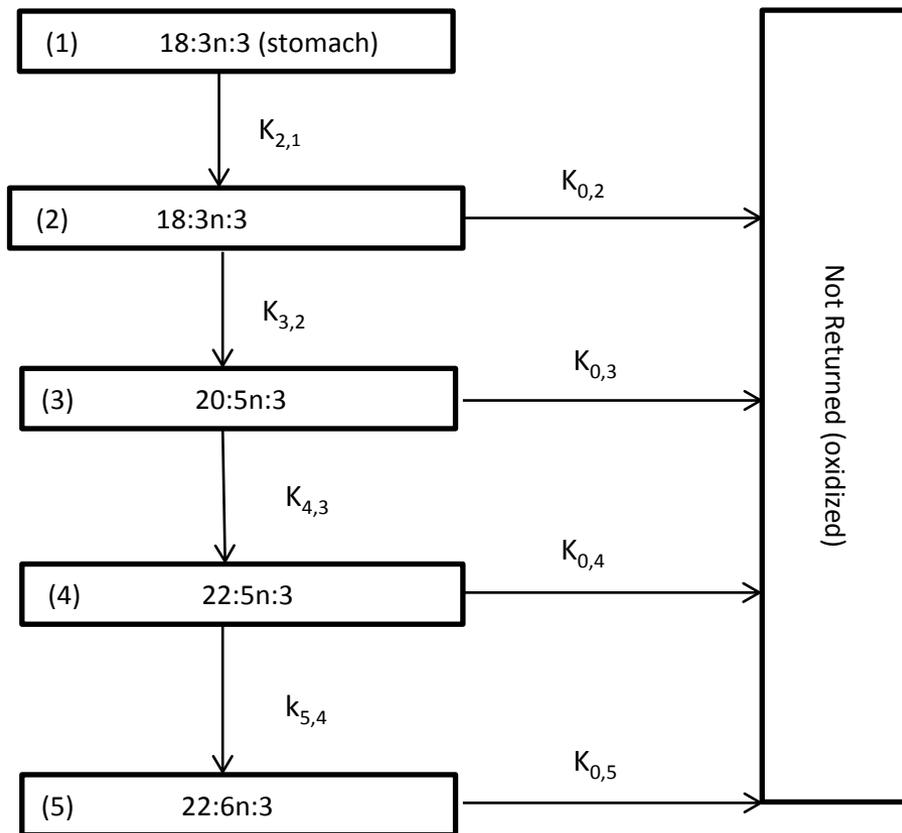


Fig 1: Compartmental model for the metabolism of  $\alpha$ -linolenic acid.

- The structure of the model consists of five compartments.
- The rectangle represents each metabolite of the fatty acid in the metabolism process.
- Compartment 1 represents administration of the isotope (1g) via the gastro-intestinal tract.
- Compartments 2 to 5 represent  $n-3$  fatty acid in the plasma following on successive steps in desaturation and elongation of the label fatty acid.
- The fractional transfer rate  $K_{ij}$  is the fraction of the substrate that is transferred from substrate compartment  $j$  to product compartment  $i$ .
- The arrows indicating  $K_{0j}$  is the fraction of the substrate that is irreversibly lost by each compartment  $j$ .

## The system of differential equations

$$\frac{dc_5}{dt} = k_{5,4}c_4 - k_{0,5}c_5 \quad (1)$$

$$\frac{dc_4}{dt} = k_{4,3}c_3 - k_{0,4}c_4 - k_{5,4}c_4 \quad (2)$$

$$\frac{dc_3}{dt} = k_{3,2}c_2 - k_{0,3}c_3 - k_{4,3}c_3 \quad (3)$$

$$\frac{dc_2}{dt} = k_{2,1}c_1 - k_{0,2}c_2 - k_{3,2}c_2 \quad (4)$$

$$\frac{dc_1}{dt} = -k_{21}c_1 \quad (5)$$

The parameters  $k_{5,4}$ ,  $k_{4,3}$ ,  $k_{3,2}$ ,  $k_{2,1}$ ,  $k_{0,5}$ ,  $k_{0,4}$ ,  $k_{0,3}$  and  $k_{0,2}$  of the differential equations are to be estimated by means of linear least squares method using measured data.

## *Concentration assumption in the stomach*

*Assuming the concentration of 18:3n-6 in stomach follows a decreasing quadratic function wrt time:*

$$at^2 + bt + c$$

*Here the parameters  $a$  and  $b$  can be included in the inverse problem formulation and  $c$  is known from the initial condition of the concentration in the stomach.*

# Results & experimental data

## Estimated parameters

<b><i>Kinetics Parameters</i></b>	<b><i>Estimated values</i></b>
k21	0.000425
k02	0.1449
k32	0.00030
k03	0.0015
k04	0.0476
k43	0.01317
k05	0.0124
k54	0.0101

## Time series experimental data

Time	c2	c3	c4	c5
0	0.0026	0.0000	0.0001	0
8	0.6690	0.0067	0.0008	0.0001
24	0.0952	0.0082	0.0026	0.0003
48	0.0524	0.0073	0.0019	0.0010
72	0.0034	0.0054	0.0005	0.0007
96	0.0212	0.0014	0.0004	0.0007
168	0.0036	0.0007	0.0003	0.0019

*Subject 18 was used in the above experiment to determine the respective values of the kinetics parameters. The parameters obtained are close to those obtained in the reference paper [1] in which their results were calculated by means of the software WIMSAM.*

***Fatty acids composition of serum cholesteryl esters (%) from healthy subjects supplemented with hempseed and flaxseed oils for 4 weeks:  
Data obtained from School of Science***

Fatty acids	Hempseed oil		Flaxseed oil	
	0 week	4 week	0 week	4 week
<b>n-6</b>				
18:2n-6- LA	50.90	56.65	49.89	52.90
18:3n-6- GLA	0.57	1.19	0.65	0.43
20:3n-6 -DGLA	0.53	0.75	0.53	0.42
20:4n-6-AA	4.01	4.10	4.38	3.52
<b>n-3</b>				
18:3n-3-ALA	0.98	1.30	0.96	4.13
20:5n-3-EPA	1.22	1.10	1.46	1.72
22:6n-3-DHA	0.37	0.33	0.44	0.34

***Fatty acids composition of serum triglycerides (%) from healthy subjects supplemented with hempseed and flaxseed oils for 4 weeks: Data obtained from School of Science***

Fatty acids	Hempseed oil		Flaxseed oil	
	0 week	4 week	0 week	4 week
n-6				
18:2n-6	17.99	25.54	17.03	19.46
18:3n-6	0.43	1.01	0.47	0.36
20:3n-6	0.38	0.55	0.34	0.38
20:4n-6	2.20	2.33	2.12	1.85
n-3				
18:3n-3	2.05	3.76	1.71	9.66
20:5n-3	2.64	1.59	1.45	2.00
22:6n-3	1.97	1.41	1.68	1.45

*We need at least three columns of data for Hempseed oil and three columns of data for Flaxseed oil – not enough data at different time points in calculating the derivatives.*

*The number of unknowns in one equation is more than the number of available data*

*Need to use adjoint method [5] or regularisation ( current work).*

## *Time-course of fatty acids concentration ( $\mu\text{g}/\text{ml}$ ) of plasma triacylglycerol and cholesterol ester from healthy subjects*

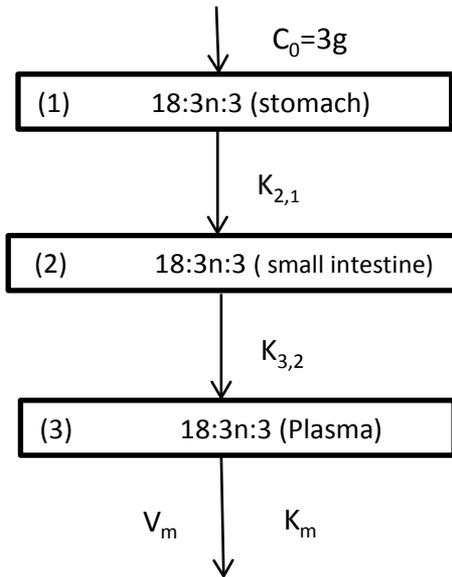
Time (h)	Triacylglycerol		Cholesterol ester	
	LA	LNA	LA	LNA
0	0	0	0	0
2	25.83	25.00	0.83	0.83
4	37.50	36.67	1.67	1.25
6	13.33	13.33	3.75	1.67
8	18.33	18.33	8.33	2.50
10	15.00	13.33	10.00	2.50
12	12.50	8.33	12.50	3.33
14	10.00	6.67	15.00	3.33
16	8.33	5.00	17.50	2.92
24	3.33	0	21.25	2.50
48	0.17	0	22.08	1.67

*The model that we are considering is a very simple metabolism model of 18:3n-3 (LNA) to 20:5n-3 (EPA) and finally to 22:6n-3 (DHA). The equations are inter-related.*

*The data provided above, say the second column, reflecting the LNA concentration at various times for 3 g of deuterium-labelled LNA intake. A graph can be easily drawn for this LNA concentration.*

*However it is possible to use the data in an absorption model [3,4]. ( current work)*

## Absorption & Elimination model



*Elimination Process (Michaelis Menten Equation)*

Where  $V_m$  and  $K_m$  M.M coefficient

- *The structure of the open model consists of Three compartments.*
- *The rectangle represents 18:3n:3 in the absorption scheme*

*The system differential equations are:*

$$\frac{dc_1}{dt} = -k_{21}c_1$$

$$\frac{dc_2}{dt} = k_{21}c_1 - k_{32}c_2$$

$$\frac{dc_3}{dt} = k_{32}c_2 - \frac{V_m c_3}{K_m + c_3}$$

## References

- [1] Pawlosky, R. J., et al. Physiological compartmental analysis of alpha-linolenic acid metabolism in adult humans. *J. Lipid Res.* 42(8):1257-1265, 2001.
- [2] G. Golub Numerical Methods for solving Linear Least Squares Problems.
- [3] J. E. PIETERS, M. WEDEL and G. SCHAAFSMA Parameter estimation in a three-compartment model for blood alcohol curves
- [4] Bus, J. C. P., Domselaar, B. V. and Kok, J. (1975) Nonlinear least squares estimation. Report NW 17/75, Mathematical Centre, Amsterdam.
- [5] S.Muller,J.Lu,P.Kugler,H.W.Engl Parameter Identification in Systems Biology: Solving Ill-posed Inverse Problems using Regularization.